

Table S4 Oligonucleotide primers used¹.

I. Primers used for qPCR studies. For primers overlapping splice sites, both involved exons are listed.

Primer	Sequence (5' to 3')	Exon
<i>egl3</i> _qP-f	GCATTCGTGCGAGGTCAAAGGC	1
<i>egl3</i> _qP-r	GCAATCCCCCATGTTCCCTTGCA	2-3
<i>ttl11</i> _qP-f	TCCCAGCATGAGGATTGAACACG	3
<i>ttl11</i> _qP-r	CCGGCATTGACGTGATGTGTTTC	4-5
<i>camk2b</i> _qP-f	AGAGGCCCGGATCTGCCGTT	1
<i>camk2b</i> _qP-r	TGCTGGTGGATGTGGCTGACG	4
<i>btr01</i> _qP-f	CGGCCATACTGTGACGGAGGC	1
<i>btr01</i> _qP-r	CTGAGACCGCAGTTCGGCGG	3
<i>ncam2</i> _qP-f	TCGGATTGCTCGTCGGTGGC	1
<i>ncam2</i> _qP-r	GCCTGGCACCGGTAGATGCC	3
<i>crtc3</i> _qP-f	GCACTCATGACAGACCTCACTGT	1
<i>crtc3</i> _qP-r	TGGAAATCAGTGCTGGCATTGCCT	2
<i>irs2</i> _qP-f	ACACAGCTCTGCCTCCGTAGA	1
<i>irs2</i> _qP-r	GGAGTAACCTCTGCTTCCTGCTCA	2
<i>sept8a</i> _qP-f	CCTCGGCGGTCATGTTGGCTT	1-2
<i>sept8a</i> _qP-r	GTTCTGGGCCGAGGTAGACG	3
<i>eef1a11</i> _qP-f	AGAAGGAAGCCGCTGAGATGG	3
<i>eef1a11</i> _qP-r	TCCGTTCTGGAGATACCAGCC	4

II. Primers used for amplification and cloning of full-length mRNAs.

Primer	Sequence (5' to 3')
<i>irs2</i> _FL-f: <u>Clal</u> / Start	GGTGGT <u>ATCGAT</u> ATGG CAAGTCCGCCTCTTA
<i>irs2</i> _FL-r: <u>XhoI</u> / Stop	GCCACC <u>CTCGAGTCA</u> ATCTTGACAGTGTTGCAG
<i>crtc3</i> _FL-f: <u>Clal</u> / Start	GGTGGT <u>ATCGAT</u> ATGT CTGGATCCCCGGGC
<i>crtc3</i> _FL-r: <u>XbaI</u> / Stop	GCCACC <u>CTAGACTA</u> AAAGTCTGCTACTGCGAAAGC
<i>btr01</i> _FL-f: <u>Clal</u> / Start	GGTGGT <u>ATCGAT</u> ATGT CATTTCCTGGTGAATTCCTGTC
<i>btr01</i> _FL-r: <u>XbaI</u> / Stop	GCCACC <u>CTAGACTA</u> TATTCTAACCAGCTTGGGTCAGC

¹ Relevant restriction sites are underlined, and start/stop codons are in bold.

III. Primers used for amplification and cloning of in situ hybridization probes. Primers containing restriction sites were directionally cloned into pCS2p+. Primers without restriction sites were TA cloned into vector pCRII and directionality was validated by sequencing.

Primer	Sequence (5' to 3')
<i>bsg</i> _IS-f: BamHI	ATGCGGATCCGGTTTCAAGCCGAAGCTATG
<i>bsg</i> _IS-r: XbaI	ATGCTCTAGAGACCCAATAGTGGCCTTTGA
<i>ythdf2</i> _IS-f: BamHI	ATGCGGATCCCAAACAACGCGCAGTCTAA
<i>ythdf2</i> _IS-r: XbaI	ATGCTCTAGACAGTGCTCCAGACTGTCCAA
<i>smarca5</i> _IS-f: BamHI	ATGCGGATCCGGAGAGCGTTTACGATGAGC
<i>smarca5</i> _IS-r: XbaI	ATGCTCTAGACGACAGCACAGCCGTAGTTA
<i>irs2</i> _IS-f:	CTTCAGTCAGCCCCACTAAC
<i>irs2</i> _IS-r:	CCTGCTTTACAACAACCGCC
<i>crtc3</i> _IS-f:	GGACGTTTCCCTCAGGCCTG
<i>crtc3</i> _IS-r:	GGTAGTGGGACAGACCCGCG
<i>btr01</i> _IS-f:	CGTGCATTTCTCCTACTGGG
<i>btr01</i> _IS-r:	GCAAACATACTGATGGTGCTG