**File S4**

**Supplemental Materials and Methods**

TEM Protocol

Samples were fixed with 2% gluteraldehyde and 2% paraformaldehyde in 0.1 NaCacodylate (460 mOsm). Salivary glands were fixed at room temperature for 2 hours, and transferred to 4$^{o}$C for overnight fixation. Fix was removed and samples washed three times with 0.1M NaCacodylate and 0.15M Sucrose (350 mOsm). Postfix incubation was carried out with 2% osmium tetraoxide in 0.1M NaCacodylate and 0.07M Sucrose (350 mOsm) for one hour. Samples were rinsed with 0.1M NaCacodylate in 0.15 M Sucrose (350 mOsm), and washed with distilled water three times. En-Bloc stain was carried out in 2% uranyl acetate aqueous solution in the dark for one hour. Following distilled water washes, samples were dehydrated through an ethanol series of 30%, 50%, 70%, 85% and 95% for 15 minutes each. Three one-hour incubations in 100% ethanol completed dehydration step. Samples were incubated in propylene oxide for two 30-minute periods, and then transferred to a 1.5:1 solution of LX112:PO for six hours. Samples were then placed in vacuum desiccantor overnight. Initial polymerization occurred at 45 $^{o}$C overnight, and then transferred to 60 $^{o}$C for an additional 24 hours. Blocks were allowed to cool and sectioned. Ultrathin sections were stained with uranyl acetate aqueous solution for 20 minutes, and lead citrate for 5 minutes.