Supporting Materials and Methods

Strain constructions

**CaLY226 (MET3p-MMS22/mms22::C.d.HIS1):** The SAT1-MET3p cassette from plasmid pFA-SAT1-MET3p was amplified using the primers oLY152 and oLY153 to generate a SAT1-MET3p-MMS22 cassette with 100 base pairs (bps) of homology to the MMS22 5’ upstream region and 100 bps of homology to the beginning of the MMS22 ORF. Primers oLY83 and oLY84 were used to amplify genomic DNA on the 5’ side of the MMS22 gene; primers oLY85 and oLY86 were used to amplify genomic DNA on the 3’ side of the MMS22 gene; primers oLY232 and oLY233 were used to amplify the C.d.HIS1 sequence from plasmid pSN52. These three fragments were fused to generate mms22::C.d.HIS1 disruption cassette. *C. albicans* SN152 was then transformed with these cassettes to generate strains CaLY8 (MMS22/mms22::C.d.HIS1) and CaLY226 (MET3p-MMS22/mms22::C.d.HIS1). Proper integration of SAT1-MET3p-MMS22 cassette was verified by genomic PCR using primers oLY87 and oLY300 as well as oLY88 and oLY301. Correct integration of mms22::C.d.HIS1 disruption cassette was confirmed with primers oLY87 plus oLY236 as well as oLY88 plus oLY237. Presence of MMS22 was verified with primers oLY89 and oLY90.

**CaLY337 (ARG4-MET3p-TOF1/tof1::C.m.LEU2):** The ARG4-MET3p cassette from plasmid pFA-ARG4-MET3p was amplified using the primers oLY312 and oLY313, and then fused with the upstream region (amplified with oLY534 and oLY535) and the beginning of the TOF1 ORF (amplified with oLY536 and oLY537) to generate an ARG4-MET3p-TOF1 cassette. Primers oLY160 and oLY161 were used to amplify genomic DNA on the 5’ side of the TOF1 gene; primers oLY162 and oLY163 were used to amplify genomic DNA on the 3’ side of the TOF1 gene; primers oLY232 and oLY233 were used to amplify the C.m.LEU2 sequence from plasmid pSN40. These three fragments were fused to generate tof1::C.m.LEU2 disruption cassette. *C. albicans* SN152 was then transformed with these cassettes to generate strains CaLY219 (TOF1/tof1::C.m.LEU2) and CaLY337 (ARG4-MET3p-TOF1/tof1::C.m.LEU2). Proper integration of ARG4-MET3p-TOF1 cassette was verified by genomic PCR using primers oLY461 and oLY538 as well as oLY462 and oLY301. Proper integration of tof1::C.m.LEU2 disruption cassette was verified with primers oLY461 and oLY366 as well as oLY462 and oLY367. Presence of TOF1 was verified with primers oLY463 and oLY464.
CaLY249 (ARG4-MET3p-CSM3/csm3::C.m.LEU2): The ARG4-MET3p cassette from plasmid pFA-ARG4-MET3p was amplified using the primers oLY312 and oLY313, and then fused with the upstream region (amplified with oLY543 and oLY544) and the beginning of the CSM3 ORF (amplified with oLY545 and oLY546) to generate a ARG4-MET3p-CSM3 cassette. Primers oLY465 and oLY466 were used to amplify genomic DNA on the 5’ side of the CSM3 gene; primers oLY467 and oLY468 were used to amplify genomic DNA on the 3’ side of the CSM3 gene; primers oLY232 and oLY233 were used to amplify the C.m.LEU2 sequence from plasmid pSN40. These three fragments were fused to generate csm3::C.m.LEU2 disruption cassette. *C. albicans* SN152 was then transformed with these cassettes to generate strains CaLY220 (CSM3/csm3::C.m.LEU2) and CaLY249 (ARG4-MET3p-CSM3/csm3::C.m.LEU2). Proper integration of ARG4-MET3p-CSM3 cassette was verified by genomic PCR using primers oLY469 and oLY538 as well as oLY470 and oLY301. Proper integration of csm3::C.m.LEU2 disruption cassette was verified with primers oLY469 and oLY366 as well as oLY470 and oLY367. Presence of CSM3 was verified with primers oLY471 and oLY472.

CaLY316 (ARG4-MET3p-MRC1/mrc1::C.m.LEU2): The ARG4-MET3p cassette from plasmid pFA-ARG4-MET3p was amplified using the primers oLY312 and oLY313 and then fused with the upstream region (amplified with oLY539 and oLY540) and the beginning of the MRC1 ORF (amplified with oLY541 and oLY542) to generate a ARG4-MET3p-MRC1 cassette. Primers oLY174 and oLY175 were used to amplify genomic DNA on the 5’ side of the MRC1 gene; primers oLY176 and oLY177 were used to amplify genomic DNA on the 3’ side of the MRC1 gene; primers oLY232 and oLY233 were used to amplify the C.m.LEU2 sequence from plasmid pSN40. These three fragments were fused to generate mrc1::C.m.LEU2 disruption cassette. *C. albicans* SN152 was then transformed with these cassettes to generate strains CaLY222 (MRC1/mrc1::C.m.LEU2) and CaLY316 (ARG4-MET3p-MRC1/mrc1::C.m.LEU2). Proper integration of ARG4-MET3p-MRC1 cassette was verified by genomic PCR using primers oLY477 and oLY538 as well as oLY478 and oLY301. Proper integration of mrc1::C.m.LEU2 disruption cassette was verified with primers oLY477 and oLY366 as well as oLY478 and oLY367. Presence of MRC1 was verified with primers oLY479 and oLY480.

CaLY235 (rad57::C.m.LEU2/rad57::C.d.ARG4): Primers oLY481 and oLY482 were used to amplify genomic DNA on the 5’ side of the RAD57 gene; primers oLY483 and oLY484 were used to amplify genomic DNA on the 3’ side of the RAD57 gene; primers oLY232 and oLY233 were used to amplify the C.m.LEU2 and C.d.ARG4 sequence from plasmid pSN40 and pSN69. These fragments were fused to generate
rad57::C.m.LEU2 and rad57::C.d.ARG4 disruption cassettes. C. albicans SN152 was then transformed with these cassettes to generate strains CaLY223 (RAD57/rad57::C.m.LEU2) and CaLY235 (rad57::C.m.LEU2/rad57::C.d.ARG4). Proper integration of rad57::C.m.LEU2 cassette was verified by genomic PCR using primers oLY485 and oLY366 as well as oLY486 and oLY367. Proper integration of rad57::C.d.ARG4 disruption cassette was verified with primers oLY485 and oLY238 as well as oLY486 and oLY239. Presence of RAD57 was verified with primers oLY487 and oLY488.

CaLY236 (rtt101::C.m.LEU2/rtt101::C.d.ARG4): Primers oLY489 and oLY490 were used to amplify genomic DNA on the 5’ side of the RTT101 gene; primers oLY491 and oLY492 were used to amplify genomic DNA on the 3’ side of the RTT101 gene; primers oLY232 and oLY233 were used to amplify the C.m.LEU2 and C.d.ARG4 sequence from plasmid pSN40 and pSN69. These fragments were fused to generate rtt101::C.m.LEU2 and rtt101::C.d.ARG4 disruption cassettes. C. albicans SN152 was then transformed with these cassettes to generate strains CaLY224 (Rtt101/rtt101::C.m.LEU2) and CaLY236 (rtt101::C.m.LEU2/rtt101::C.d.ARG4). Proper integration of rtt101::C.m.LEU2 cassette was verified by genomic PCR using primers oLY493 and oLY366 as well as oLY494 and oLY367. Proper integration of rtt101::C.d.ARG4 disruption cassette was verified with primers oLY493 and oLY238 as well as oLY494 and oLY239. Presence of Rtt101 was verified with primers oLY495 and oLY496.

CaLY251 (MET3p-MMS22/mms22::C.d.HIS1 ARG4-MET3p-MRC1/mrc1::C.m.LEU2): C. albicans CaLY226 was transformed with mrc1::C.m.LEU2 and ARG4-MET3p-MRC1 cassettes to generate strains CaLY228 (MET3p-MMS22/mms22::C.d.HIS1 MRC1/mrc1::C.m.LEU2) and CaLY251 (MET3p-MMS22/mms22::C.d.HIS1 ARG4-MET3p-MRC1/mrc1::C.m.LEU2).

CaLY246 (MET3p-MMS22/mms22::C.d.HIS1 ARG4-MET3p-CSM3/csm3::C.m.LEU2): C. albicans CaLY226 was transformed with csm3::C.m.LEU2 and ARG4-MET3p-CSM3 cassettes to generate strains CaLY234 (MET3p-MMS22/mms22::C.d.HIS1 CSM3/csm3::C.m.LEU2) and CaLY246 (MET3p-MMS22/mms22::C.d.HIS1 ARG4-MET3p-CSM3/csm3::C.m.LEU2).

CaLY242 (MET3p-MMS22/mms22::C.d.HIS1 rad57::C.m.LEU2/rad57::C.d.ARG4): C. albicans CaLY226 was transformed with rad57::C.m.LEU2 and rad57::C.d.ARG4 cassettes to generate strains CaLY238 (MET3p-MMS22/mms22::C.d.HIS1 RAD57/rad57::C.m.LEU2) and CaLY242 (MET3p-MMS22/mms22::C.d.HIS1
rad57::C.m.LEU2/rad57::C.d.ARG4).

**CaLY244** (MET3p-MMS22/mms22::C.d.HIS1 rtt101::C.m.LEU2/rtt101::C.d.ARG4): *C. albicans* CaLY226 was transformed with rtt101::C.m.LEU2 and rtt101::C.d.ARG4 cassettes to generate strains CaLY240 (MET3p-MMS22/mms22::C.d.HIS1 Rtt101/rtt101::C.m.LEU2) and CaLY244 (MET3p-MMS22/mms22::C.d.HIS1 rtt101::C.m.LEU2/rtt101::C.d.ARG4).