Endoplasmic reticulum stress-related genes in yellow catfish *Pelteobagrus fulvidraco*: molecular characterization, tissue expression and expression responses to dietary copper deficiency and excess

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Running title: Endoplasmic reticulum stress-related genes in fish

Keywords: Pelteobagrus fulvidraco; molecular characterization; Dietary Cu; ER stress; unfolded protein response

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ABSTRACT

Two endoplasmic reticulum (ER) molecular chaperones [glucose-regulated protein 78 (GRP78) and calreticulin (CRT)] and three ER stress sensors [PKR-like ER kinase (PERK), inositol requiring enzyme (IRE)-1α, and activating transcription factor (ATF)-6α] cDNAs were firstly characterized from yellow catfish, *Pelteobagrus fulvidraco*. The predicted amino acid sequences for the yellow catfish GRP78, CRT, PERK, IRE-1α and ATF-6α revealed that the proteins contained all of the structural features that were characteristic of the five genes in other species, including the KDEL motif, signal peptide, sensor domain and effector domain. mRNAs of the five genes mentioned above were expressed in various tissues, but their mRNA levels varied among tissues. Dietary Cu excess, but not Cu deficiency, activated the chaperones (GRP78 and CRT) and folding sensors in ER, and the UPR signaling pathways (i.e., PERK–eIF2α and the IRE1–XBP1) in a tissue-specific manner. For the first time, our study cloned GRP78, CRT, PERK, IRE-1α and ATF-6α genes in yellow catfish and demonstrated their differential expression among tissues. Moreover, the present study also indicated differential regulation of these ER stress-related genes by dietary Cu deficiency and excess, which will be beneficial for us to evaluate effects of dietary Cu levels in fish at the molecular level, based on the upstream pathway of lipid metabolism—the ER, and thus provide novel insights on the nutrition of Cu in fish.
The endoplasmic reticulum (ER) is an important intracellular organelle responsible for protein and lipid synthesis, xenobiotic detoxification and cellular calcium storage (Borgese et al., 2006; Gorlach et al., 2006). Conditions disrupting the ER homeostasis cause accumulation of unfolded and misfolded proteins in the ER lumen, create a state defined as ER stress, and activate a complex signaling network termed unfolded protein response (UPR) (Schröder and Kaufman, 2005). UPR is known to trigger a unique signaling cascade from the ER to the nucleus, being characterized by enhanced expression of ER-resident chaperones, such as glucose-regulated protein 78 (GRP78) (Kozutsumi et al., 1988), and calreticulin (CRT) (Caspersen et al., 2000). In addition, UPR is characterized by downregulation of protein translation through three ER-transmembrane transducers: PKR-like ER kinase (PERK), inositol requiring enzyme (IRE)-1α, and activating transcription factor (ATF)-6α (Harding et al., 1999). In the unstressed state, the ER lumenal domains of all three stress sensors are bound by the ER chaperone GRP78. Upon ER stress, GRP78 binding to unfolded proteins causes dissociation from three ER-transmembrane transducers, leading to their activation (Bertolotti et al., 2000; Oikawa et al., 2009). Following activation, UPR signaling pathways act to induce expression of genes that encode functions to ameliorate the stressed state of the ER.

During the last few years, ER stress and UPR-related proteins have been extensively analyzed at the molecular level in mammals. Completely characterized cDNA and genomic sequences of GRP78, CRT, PERK, IRE-1α and ATF-6α have been previously reported in human, mouse and rat species (Schröder and Kaufman, 2005). However, in fish, to our knowledge, full-length cDNA sequences of the ER stress-related genes (GRP78, CRT, PERK, IRE-1α and ATF-6α) were only determined in very limited fish species, such as grass carp Ctenopharyngodon idella (GRP78, Zhu et al., 2013), rainbow trout Oncorhynchus
mykiss (CRT, Kales et al., 2004), Asian seabass Lates calcarifer (CRT, Bai et al., 2012), and medaka Oryzias latipes (ATF6, Ishikawa et al., 2011), although some partial sequences have been reported for other fish species (Ojima et al., 2005; Komoike and Matsuoka 2013; Chen et al. 2014; Das et al. 2015; Li and Li, 2015). Although their expression pattern was often covered in these papers, study involved in their physiological function was often absent. Thus, identification of ER stress-related genes in other teleosts was a key step for characterizing the role and mechanism of ER stress.

Copper (Cu) is an essential micronutrient for vertebrate animals including fish (Watanabe et al., 1997). It has numerous functions in cellular biochemistry including vital roles in cellular respiration, and as a co-factor for over 30 different enzymes (Watanabe et al., 1997). At present, optimal dietary Cu requirement has been determined in many fish species, ranging from 3 to 10 mg Cu kg⁻¹ feed, which depends on the species, feeding regime and life stage (NRC, 2011). Studies have also shown that over-loading of dietary Cu in fish caused toxic syndrome (Berntssen et al., 1999; Lundebye et al., 1999; Shiau and Ning, 2003). In contrast, dietary Cu deficiency has been shown to reduce appetite, growth and cause anemia in several fish (Gatlin and Wilson, 1986; Shiau and Ning, 2003; Lin et al., 2008; Tan et al., 2011). Recently, our own studies have pointed out that dietary Cu deficiency and excess could result in the changes of lipid deposition and metabolism in fish (Chen et al., 2015). However, the molecular mechanisms of the upstream pathway of lipid metabolism underlying dietary Cu-induced alteration in lipid metabolism have not been elucidated. Studies have pointed out that ER stress is one of the cellular stresses reported to induce lipid accumulation (Lee et al., 2008) and ER stress played crucial roles in hepatic lipid metabolism in mammals (Sriburi et al., 2004; Rutkowski et al., 2008; Kammoun et al., 2009). Thus, considering the important role ER plays in lipid metabolism, we hypothesize that ER stress-dependent alteration in lipid
homeostasis was the mechanism that underlies the change of lipid deposition of yellow catfish in responses to dietary copper levels. Yellow catfish *Pelteobagrus fulvidraco*, an omnivorous freshwater fish, is widely distributed in the inland freshwater rivers in China. The fish is regarded as a potential model for studying lipid metabolism in fish because it has relatively high lipid accumulation under natural conditions (Zheng et al., 2013a). The cDNA sequences and molecular characterization of many genes have been elucidated in the fish species in our laboratory (Gong et al., 2013; Zheng et al., 2013a, b, Chen et al., 2014; Song et al., 2014; 2015a, b). Recent studies in our laboratory indicated that dietary Cu deficiency and excess could influence lipid deposition and metabolism in *P. fulvidraco* (Chen et al., 2015). However, these studies only determined the change of lipid metabolism-related genes expression and enzyme activities after dietary Cu treatment. The molecular and regulatory mechanisms in the upstream pathway of lipid metabolism under dietary Cu treatment have not been explored. Clearly, an understanding of the molecular basis of ER stress could underpin efforts to address this problem. Therefore, in this study, the full-length cDNA sequences of two ER molecular chaperones (GRP78 and CRT) and three ER stress sensors (PERK, IRE-1α and ATF-6α) genes were first cloned and their tissue specific expressions were determined. Meanwhile, effect of dietary Cu deficiency and excess on their mRNA levels was explored. The results will be beneficial for us to elucidate the molecular basis of ER stress, and to further evaluate effects of dietary Cu levels in fish at a molecular level, based on the upstream pathway of lipid metabolism, which will greatly extend our understanding of the nutrition of Cu in fish.

**MATERIALS AND METHODS**
Here, two experiments were conducted. The first experiment was involved in the cloning and tissue expression profile analysis of GRP78, CRT, PERK, IRE-1α and ATF-6α transcripts. The second experiment was designed to evaluate a possible transcriptional regulation of GRP78, CRT, PERK, eIF2α, IRE-1α, XBP-1 and ATF-6α by dietary Cu deficiency and excess. We ensured that the experiments performed on animals followed the ethical guidelines of Huazhong Agricultural University and Animal Care and Use Committee (IACUC) in Huazhong Agricultural University specifically approved this study.

Experiment 1: GRP78, CRT, PERK, IRE-1α and ATF-6α cDNAs cloning and mRNA expression patterns of various tissues

Experimental animals: Yellow catfish (1.83 ± 0.05 g, mean ± SEM) for cloning the GRP78, CRT, PERK, IRE-1α and ATF-6α genes were obtained from a local fish pond (Wuhan, China) and transferred to indoor fiberglass tanks (400-l in water volume) for two-week acclimation. During the acclimation period, the fish were fed to apparent satiation twice daily with a commercial pellet diet. The experiment was conducted at ambient temperature and subjected to natural photoperiod (approximately 14 h light/10 h darkness). Water quality parameters were monitored twice a week in the morning. The parameters were followed: water temperature, 27.9 ± 1.2 °C; pH, 7.66 ± 0.11; dissolved oxygen, 6.34 ± 0.15 mg l⁻¹; total hardness, 151.8 ± 1.1 mg l⁻¹ as CaCO₃. Fish were anesthetized with 3-aminobenzoic acid ethyl ester methanesulfonate (MS-222 at 10 mg l⁻¹) and sacrificed by decapitation. Skeletal muscle, gill, head kidney, mid-intestine, brain, spleen, liver, heart, testis and ovary form six fish were dissected and immediately prepared for RNA isolation (n=6).
Each sample was pulverized with its own mortar and pestle, and the mortars and pestles were heated at 230 °C for 8h to inactivate RNases. Total RNA was extracted using TRIzol RNA reagent (Invitrogen, USA) based on the acid guanidinium thiocyanate-phenol-chloroform extraction method, according to manufacturer’s instructions. The quality of total RNA was checked by agarose gel electrophoresis. The concentration and the ratio (between 1.8 and 2.0) of OD\textsubscript{260}/OD\textsubscript{280} of total RNA were determined using a Nanodrop ND-2000 spectrophotometer (Thermo Fisher Scientific, USA). Total RNA (1 μg) was reverse-transcribed to cDNA with oligo-dT primers and a cDNA Synthesis Kit (TaKaRa, Japan), according to manufacturer’s instructions.

**Cloning and sequencing of GRP78, CRT, PERK, IRE-1α and ATF-6α cDNAs:** In the present study, we used the liver of yellow catfish to clone the cDNA sequences of the five genes. Degenerate primers (Table 1) designed based on the most conserved regions of the fish GRP78, CRT, PERK, IRE-1α and ATF-6α sequences available in the GenBank and Ensembl databases, were used to amplify partial cDNA fragments of GRP78, CRT, PERK, IRE-1α and ATF-6α. The PCR program consisted of initial denaturing for 4 min at 94 °C followed by 30 cycles of 94 °C for 30 s, 55 °C for 30 s, 72 °C for 1 min and a final elongation step at 72 °C for 5 min, with 500 ng cDNA template, 0.2 μM primer, 5 μl 10 × Ex Taq Buffer, 2.5mM dNTP, and 1.25 U Ex Taq polymerase (TaKaRa, Japan). The target fragments were purified using the EZNA gel extraction kit (Omega, USA) according to manufacturer’s instructions, and then sub-cloned into the pMD 19-T cloning vector (TaKaRa, Japan). Positive clones containing inserts of an expected size were sequenced, with sanger dideoxy mediated chain termination method (Sangon, China). In order to obtain the 3’ and 5’ end
sequences of GRP78, CRT, PERK, IRE-1α and ATF-6α, nested 3’ and 5’ RACE PCR were performed with a SMART RACE cDNA Amplification Kit (Clontech, USA) using the DNase treated total RNA, with 1 μl 10 × Reaction Buffer, 1 U DNase I for 1 μg RNA incubated at 37 °C for 15 min, then with 1 μl 25 mM EDTA incubated at 65 °C for 15 min to inactivate the DNase I. In the first PCR, the cDNA was amplified with two outer primer sets (Table 1) and Universal Primer Mix (provided in the kit) using Advantage® 2 PCR Kit (Clontech, USA), with 200 ng cDNA template, 10 μM primer, 4 μl 10 × LA PCR Buffer, 25 mM MgCl₂, and 1.25 U LA Taq (TaKaRa, Japan). In the second PCR, inner primer sets (Table 1) and Nested Universal Primer (provided in the kit) were used, with 1 μl 1st cDNA product, 2.5 mM dNTP, 10 μM primer, 5 μl 10 × LA PCR Buffer, 25 mM MgCl₂, and 2.5 U LA Taq (TaKaRa, Japan). The PCR parameters were 30 cycles at 94 °C for 30 s, 55 °C for 30 s, and 72 °C for 1 min, with an additional initial 3-min denaturation at 94 °C and a 10-min final extension at 72 °C. The RACE products were sequenced as described above.

**Sequence analysis:** The core fragment, 3’ end and 5’ end sequences were assembled using SeqMan II software in DNASTAR PACKAGE to obtain full-length GRP78, CRT, PERK, IRE-1α and ATF-6α cDNAs. The sequences were edited and analyzed using the program EDITSEQ of DNASTAR package to search for the open reading frame (ORF) and then translated into amino acid sequences using standard genetic codes. The nucleotide sequences were compared with DNA sequences present in the GenBank database using BLAST network service at the NCBI (http://blast.ncbi.nlm.nih.gov/). Sequence alignments and percentage of amino acid conservation were assessed with the Clustal-W multiple alignment algorithm. For phylogenetic analysis, multiple sequence alignments were made with
MAFFT (Katoh et al., 2002) using an amino acid model at the GUIDANCE web-server (http://guidance.tau.ac.il/) (Penn et al., 2010), which pruned unreliably aligned regions by rejecting columns with confidence scores below 0.93. The phylogenetic tree was constructed with MEGA 5.0 (Tamura et al., 2011) by the neighbor-joining (NJ) method based on the JTT+G model (Jones et al., 1992), the best-fit model of sequence evolution obtained by ML model selection. The confidence of each node was assessed by 1,000 bootstrap replicates.

**Experiment 2: Responses of mRNA Expression of GRP78, CRT, PERK, eIF2α, IRE-1α, XBP-1 and ATF-6α to dietary Cu deficiency and excess**

**Diet preparation:** Three experimental diets, in agreement with the study by Chen et al. (2015), were formulated with CuSO₄·5H₂O supplemented at levels of 0, 0.013 and 0.39 g kg⁻¹ diet at the expense of cellulose (Table 2). Different Cu contents were added to the diets, based on our recent study (Tan et al., 2011), in order to produce three different dietary Cu groups (Cu deficiency, adequate Cu and Cu excess, respectively). The formulation of the experimental diets was detailed in Chen et al. (2015). The formulated diets were dried at 80 °C in an oven until the moisture was reduced to less than 10%. The dry pellets were placed in plastic bags and stored at −20 °C until fed. Final Cu concentrations in the experimental diets were analyzed in triplicate using graphite furnace atomic absorption spectrometry (GFAAS, Chen et al., 2015), and the contents were 0.76 (Cu deficiency), 4.18 (adequate Cu), and 92.45 (Cu excess) mg Cu kg⁻¹ diet, respectively.

**Experimental procedures:** Experimental procedures are in agreement with these in our paralleled study (Chen et al. 2015). Briefly, yellow catfish were obtained from a local fish pond (Wuhan, China) and transferred to indoor fiberglass tanks (400 l in water volume) for two-week
acclimation. At the beginning of the experiment, 30 uniform-sized fish (1.83 ± 0.05 g, mean ± SEM) were stocked in each fiberglass tank (the loading of fish was about 0.183 g l⁻¹) and subjected to natural photoperiod (approximately 14 h light/10 h darkness). Each diet was assigned to three tanks in a completely randomized design, with 9 tanks for the experiment. Water in each tank was renewed 80% twice daily, before feeding. They were provided with continuous aeration to maintain the dissolved oxygen level near saturation. The fish were fed to apparent satiation twice daily at two equal meals (09:00 and 16:30 h) during the week. Care was taken to ensure that no non-eaten food remained in the tanks during feeding, thus leaching of CuSO₄ into water was very low and negligible. Cu concentrations in water samples collected 10 min before and after feeding remained low throughout the experiment (1.7 ± 0.2 μg 1⁻¹). The experiment continued for 8 weeks. At the end of the 8-wk period, 24 h after the last feeding, all fish were euthanized with MS-222 (10 mg l⁻¹). Three fish per tank were randomly selected. After the skin was rapidly removed from muscle, the liver and skeletal muscle above the lateral line were removed immediately using sterile forceps, frozen in liquid nitrogen, and stored at −80 °C (not longer than 2 weeks) for total RNA extraction (n = 3 replicate tanks, and three fish were sampled for each tank).

Quantitative real-time PCR (qPCR):
Analyses on the distribution and mRNA expression levels of GRP78, CRT, PERK, eIF2α, IRE-1α, XBP-1 and ATF-6α were conducted by quantitative real-time PCR (qPCR) method. Primers used for qPCR were shown in Table 3. Total RNA extraction and first strand cDNA synthesis were performed according to previously described methods (Chen et al., 2014). The cDNA synthesis reactions were diluted to 200 μl in water. Real-time PCR was performed in a 20 μl reaction mixture including 10 μl SYBR Premix Ex TaqTM
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II (2 ×), 0.4 μl each primer (25 μM), 1μl
diluted first-strand cDNA (5 ng) product
and 8.2 μl ddH2O. Reactions were based
on a three-step method as followed: at
95 °C for 30 s, 40 cycles of 5 s at 95 °C,
10 s at 57 °C and 30 s at 72 °C. All
reactions were performed in duplicate and
each reaction was verified to contain a
single product of the correct size by
agarose gel electrophoresis. A
non-template control and dissociation
curve were performed to ensure that only
one PCR product was amplified and that
stock solutions were not contaminated. A
10-fold dilution series was created from a
random pool of cDNA from our sample
groups ranging from × 1 dilution to ×
100,000 dilutions. The PCR efficiency and
correlation coefficients of each primer pair
were generated using the slopes of the
standard curves. The amplification
efficiencies of all genes were
approximately equal and ranged from 98
to 103 %. As housekeeping gene
sequences were not available in P. fulvidraco, a set of eight housekeeping
genres (β-actin, GAPDH, EF1A, 18S
rRNA, HPRT, B2M, TUBA and RPL17)
were selected from the literature (Zhao et
al., 2011) in order to test their transcription
stability. The relative expression levels
were calculated using the 2−ΔΔCt method
(Livak and Schmittgen, 2001) when
normalizing to the geometric mean of the
best combination of two genes (β-actin
and GAPDH, M=0.27) as suggested by
genNorm (Vandesompele et al., 2002). Prior
to the analysis, experiments were
performed to check the stability of
housekeeping genes, from which β-actin
and GAPDH showed the most stable level
of expression across the experimental
conditions (Chen et al., 2014).

Statistical analysis

Quantitative data were expressed as means
± SEM. Prior to statistical analysis, all
data were tested for normality of
distribution using the
Kolmogorov–Smirnov test. The
homogeneity of variances among the
different tissues was tested using the Barlett’s test. Data were then subjected to one-way ANOVA and Tukey’s multiple range tests. Differences between control (adequate Cu) and individual Cu-treated group (Cu deficiency and Cu excess) were analyzed by Student's T-test for independent samples. Difference was considered significant at P < 0.05. All statistics were performed using the SPSS 17.0 for Windows (SPSS, Michigan Avenue, Chicago, IL, USA).

RESULTS

Molecular characterization of GRP78, CRT, PERK, IRE-1α and ATF-6α cDNAs

In the present study, by RT-PCR, 3’- and 5’-RACE PCR, we obtained the full-length cDNA sequences of GRP78, CRT, PERK, IRE-1α and ATF-6α genes, which were shown to be 2544 bp, 1678 bp, 3776bp, 3440 bp and 2785 bp, respectively (Table 4). Sequencing revealed that their cDNAs included an ORF of 1956 bp, 1254 bp, 3240 bp, 3113 bp and 1971 bp, respectively, encoding a protein of 652 amino acids, 418 amino acids, 1080 amino acids, 1038 amino acids and 657 amino acids, respectively. The protein sequence of P. fulvidraco GRP78 possessed all the characteristic features of GRP78, including signal peptide, C-terminal KDEL motif, ATPase domain and substrate-binding domain (Fig. 1). The protein sequence of yellow catfish CRT possessed all the characteristic features of CRT, including signal peptide, N-glycosylation site and C-terminal KDEL motif (Fig. 2). The predicted amino acid sequence for the yellow catfish PERK, IRE-1α and ATF-6α possessed all the characteristic features of ER stress-sensing genes, including signal peptide, transmembrane domain, protein kinase, ribonuclease, sensor domain and effector domain (Fig. 3, 4 and 5).

Alignment of predicted polypeptide sequences showed that the amino acid sequences of yellow catfish GRP78, CRT,
PERK, IRE-1α and ATF-6α were similar to those from other fish and mammals, exhibiting 91.9–94.9%, 69.9–84.8%, 56.8–74.4%, 65.1–81.7% and 40.2–65.0% amino acid sequence identities, respectively (Table 5).

The phylogenetic analysis showed that all teleost GRP78 formed an independent cluster, while amphibian and mammalian GRP78 formed another cluster. The *P. fulvidraco* GRP78 was closer with *Astyanax mexicanus* and more distant from *Oryzias latipes* (Fig. 6A). Yellow catfish CRT clustered most closely with *Astyanax mexicanus* and *Danio rerio*, and more distantly from *Oryzias latipes* (Fig. 6B). Based on the phylogentic analysis, all teleost ER stress-sensing genes (PERK, IRE-1α and ATF-6α) formed an independent cluster, while amphibian and mammalian corresponding proteins formed another cluster, clustered the yellow catfish most closely with *Astyanax mexicanus* and *Danio rerio*, and more distantly from *Oryzias latipes* (Fig. 6C, D).

mRNA tissue expression of GRP78, CRT, PERK, IRE-1α and ATF-6α

The GRP78, CRT, PERK, IRE-1α and ATF-6α genes from *P. fulvidraco* were detected in all sampled tissues, but their mRNA abundance varied (Fig. 7). GRP78 mRNA levels were the highest in liver, followed by heart, spleen, brain, head kidney, mid-intestine, and ovary, and lowest in testis, gill and skeletal muscle. CRT mRNA levels were the highest in liver, followed by brain, heart, spleen, testis and head kidney, and the lowest in ovary, mid-intestine, gill and skeletal muscle. PERK mRNA levels were the highest in liver and heart, followed by spleen, brain, head kidney, ovary, testis and mid-intestine, and the lowest in skeletal muscle and gill. IRE-1α mRNA expression is the highest in the liver, followed by heart, spleen, mid-intestine, brain, head kidney and testis, and the lowest in skeletal muscle and gill. ATF-6α mRNA levels were the highest in liver and
heart, followed by brain, spleen, head kidney, mid-intestine and gill, and the lowest in ovary, testis and skeletal muscle.

**Effect of dietary Cu deficiency and excess on expression of GRP78, CRT, PERK, eIF2α, IRE-1α, XBP-1 and ATF-6α**

The effect of dietary Cu deficiency and excess on expression of GRP78, CRT, PERK, eIF2α, IRE-1α, XBP-1 and ATF-6α mRNAs in the liver and muscle of *P. fulvidraco* was shown in Fig. 8. In liver, compared to dietary adequate Cu group, dietary Cu excess significantly upregulated mRNA expression levels of GRP78, CRT, PERK, eIF2α, IRE-1α and XBP-1, and dietary Cu deficiency only significantly upregulated mRNA expression levels of CRT but showed no significant effects on other tested genes’ expression. In muscle, compared to dietary adequate Cu group, dietary Cu excess significantly upregulated mRNA levels of GRP78, CRT and XBP-1, but showed no significant effects on mRNA expression of PERK, eIF2α, IRE-1α, XBP-1 and ATF-6α. mRNA levels of all seven tested genes showed no significant differences between dietary Cu deficiency group and dietary adequate Cu groups in skeletal muscle.

**DISCUSSION**

To date, the five full-length GRP78, CRT, PERK, IRE-1α and ATF-6α cDNAs have only been cloned in limited fish species, such as grass carp *Ctenopharyngodon idella* (GRP78, Zhu et al., 2013), rainbow trout *Oncorhynchus mykiss* (CRT, Kales et al., 2004), Asian seabass *Lates calcarifer* (CRT, Bai et al., 2012), and medaka *Oryzias latipes* (ATF6, Ishikawa et al., 2011). In this study, for the first time, we identified the full-length cDNA sequence of all the five genes and explored their tissue distribution profiles from yellow catfish, and also indicated that the majority of their mRNA expression levels were differentially regulated by dietary Cu deficiency and excess, which would...
contribute to our understanding on the molecular basis of ER stress and would also greatly extend our understanding of the nutrition of Cu in fish.

In the present study, the ORF amino acid sequences of yellow catfish GRP78, CRT, PERK, IRE-1α and ATF-6α shared the high identity in the range of 91.9–94.9%, 69.9–84.8%, 56.8–74.4%, 65.1–81.7% and 40.2–65.0% to those of other species, in agreement with other reports (Harding et al., 1999; Higashino et al., 2006; Ishikawa et al., 2011; Lenartowski et al., 2014; Das et al., 2015). The predicted amino acid sequences for the yellow catfish ER molecular chaperones (GRP78 and CRT) revealed that the peptide contained all of the structural features that are characteristic of ER molecular chaperones in other species, containing 16 and 18-residue signal peptide sequences and the KDEL motif (Kales et al., 2004; Higashino et al., 2006; Bai et al., 2012; Das et al., 2015). The signal peptide at the N terminus and the consensus tetrapeptide KDEL at the C terminus of the amino acid sequence, ensure the protein targeting to the ER. KDEL is a retrieval motif essential for the precise sorting of ER resident proteins along the secretory pathway. It is also known that GRP78 has two major domains, a N-terminal domain containing the ATPase catalytic site and a C-terminal substrate-binding domain (Gaut and Hendershot, 1993; Gething, 1999; Das et al. 2015), which are also conserved in the GRP78 of yellow catfish. Structural predictions of CRT indicated that the protein had at least three domains, i.e. N, P, and C domains. The P-domain of CRT comprises a proline-rich sequence with three repeats of the amino acid sequence PXXIXDPAXKPEDWDE followed by three repeats of the sequence GXWXPPXIXNPXYX. Similar results were also obtained in Japanese monkey (Macaca fuscata) (Higashino et al., 2006). The P-domain binds Ca$^{2+}$ with high affinity, and the repeats may be essential
for the high affinity of Ca\(^{2+}\) binding of
CRT (Fliegel et al., 1989; Ellgaard et al.,
2001). The predicted amino acid sequence
of the yellow catfish PERK revealed that
the peptide contained the structural
features that are characteristic of PERK,
such as a signal peptide, a transmembrane
domain, a cytosolic Ser/Thr kinase domain,
sensor domain and effector domain (Cui et
al., 2011). IRE-1\(\alpha\) from mammals are
characterized by an amino-terminal ER
lumenal domain, a transmembrane region,
and a carboxy-terminal domain that
contains both kinase and RNase catalytic
activities (Ishikawa et al., 2011), which
are also conserved in the IRE-1\(\alpha\) of yellow
catfish. The predicted amino acid
sequence of ATF-6\(\alpha\) from yellow catfish
and other species are characterized by a
cytoplasmic N-terminus that contains a
bZIP motif, which functions as a
transcription factor following regulated
intramembrane proteolysis in ER-stressed
cells (Ishikawa et al., 2011). Phylogenetic
analysis suggested that yellow catfish
peptides were closely related to other fish
with similar taxonomic classes, similar to
those reported in other fish (Kales et al.,
2004).

The analysis of tissue distribution
profiles for genes is useful to better
understand their physiological roles. At
present, in fish several studies have been
involved in mRNA expression of GRP78
(Hori et al., 2010; Zhu et al., 2013; Rebl et
al., 2014; Das et al. 2015) and CRT (Kales
et al., 2004; Bai et al., 2012), and other
reports in mRNA expression profiles of
ER stress-relevant transcripts in fish (Chen
et al., 2014; Komoike and Matsuoka,
2013). However, the mRNA expression
profiles of PERK, IRE-1\(\alpha\) and ATF-6\(\alpha\)
were still absent in fish. The present study
indicated that GRP78 mRNA levels were
the highest in liver, followed by heart and
brain. The non-induced level of expression
of GRP78 indicated that it might play
multiple functions in yellow catfish as its
homologues reported in other living
organisms (Xi and Ma, 2013). Zhu et al. (2013) indicated that the expression of GRP78 in the liver of grass carp, *Ctenopharyngodon idella*, was higher than other tissues. Other studies also indicated higher GRP78 accumulation in brain and heart tissue in mammals (Higashino et al., 2006; Xi and Ma, 2013) and other tissues. The present study indicated CRT mRNA levels in testis. CRT has been identified in the sperm of several mammals (Ho and Suarez, 2003; Naaby-Hansen et al., 2001), and appears to be involved in regulating sperm capacitation, motility, and acrosome reaction (Naaby-Hansen et al., 2001; Ho and Suarez, 2003). In Asian seabass, Bai et al. (2012) observed that the highest expression was seen in the liver followed by that in the brain, spleen, and kidney. In Japanese monkey, Higashino et al. (2006) pointed out that CRT mRNA was expressed ubiquitously in every tissue except a low expression in the thymus and skeletal muscle. The present study also detected the low CRT mRNA expression in the intestine, in agreement with the reports by Kales et al. (2004). The present study indicated the CRT mRNA levels in testis. CRT has been identified in the sperm of several mammals (Ho and Suarez, 2003; Naaby-Hansen et al., 2001), and appears to be involved in regulating sperm capacitation, motility, and acrosome reaction (Naaby-Hansen et al., 2001; Ho and Suarez, 2003). Like mammals (Koyabu et al., 1994) and rainbow trout (Kales et al., 2007), yellow catfish CRT may play only a minor role in skeletal muscle, conserving its role in calcium modulation for smooth muscle such as the heart (Waisman et al., 1986). The present study reported the ubiquitous mRNA expression of the three ER stress sensors (PERK, IRE-1α, and ATF-6α) in all tested tissues with the highest expression in liver, and the lowest in skeletal muscle and gill. At present, mRNA expression profiles of these genes in fish were very scarce. Studies pointed out that the expression level of ER stress proteins was expected to
be high in secretory cells, as these cells are rich in ER (Palade, 1975). Their commonly high expression in the liver, spleen and head kidney is consistent with this hypothesis. The present study also indicated the high expression of ER stress-related genes was found in the heart. Studies suggested that ER stress might protect the heart, and even foster hypertrophic growth of the myocardium in mammals (Okada et al., 2004; Mao et al., 2006).

In mammals, lipid droplet accumulation is often induced by ER stress (Ozcan et al., 2004; Oyadomari et al., 2008). Studies indicated Cu could act as a modifier in lipid metabolism (Burkhead and Lutsenko, 2013) and the variations in dietary Cu have a profound effect on lipid deposition in fish (Berntssen et al., 1999; Tan et al., 2011; Shaw and Handy, 2006). Recently, we have indicated that dietary deficiency and excess could influence the key lipid metabolism-related genes expression (i.e., 6PGD, G6PD, FAS, ACCα, PPARγ, LXR, HSL, PPARα and ATGL) and enzyme activities (i.e., 6PGD, G6PD and FAS) in yellow catfish (Chen et al., 2015). In order to shed light on the molecular mechanisms underlying the change of lipid storage in yellow catfish, we analyzed the changes of mRNA expression of these ER-related genes known to play an important molecular role in ER stress-induced changes of lipid deposition. The present study, for the first time, found dietary Cu excess activated ER stress and the UPR signaling in a tissue-specific manner in yellow catfish. First, the chaperones and folding sensors in ER, including GRP78 and CRT, were significantly upregulated in liver and skeletal muscle of P. fulvidraco in dietary Cu excess groups. Up-regulation of GRP78 and CRT often act as an index of ER stress (Lee, 2005). Studies reported marked up-regulation of ER chaperones such as GRP78 and CRT in response to metal ions (Liu et al., 2006; Shinkai et al., 2010; Zhu et al., 2013).
Second, we found that two UPR branches, the PERK–eIF2α pathway and the IRE1–XBP1 pathway, were activated by dietary Cu excess in liver of *P. fulvidraco*. Studies also suggested that under ER stress, UPR pathways, i.e., PERK–eIF2α and the IRE1–XBP1, are activated to reduce the ER load of the unfolded proteins (Ron and Walter, 2007). In LLC-PK1 cells, Liu et al. (2006) found eIF2α mRNA levels were elevated in response to CdCl₂ exposure. The activation of UPR pathways will in turn influence lipid metabolism since these pathways controls hepatic lipogenesis, as suggested by several studies (Lee et al., 2008; Oyadomari et al., 2008; Lee and Glimcher, 2009). ATF6 also plays an important role in regulating hepatic lipid homeostasis (Yamamoto et al., 2007; Rutkowski et al., 2008). In the present study, dietary Cu excess did not significantly influenced ATF6α expression, which showed that the ATF6α pathway was not the main branch for the Cu-induced ER stress. Considering the lack of change in ER stress-related genes’ expression in skeletal muscle following dietary Cu treatments, we believe this lack of response in these genes’ expression levels suggests a different mode of regulation or function in fish skeletal muscle. Because the ER stress plays important roles in lipid homeostasis, the ER stress and UPR may be the fundamental reaction to the dietary Cu exposure. Thus, ER stress may be the unrecognized mechanism underlying the changes of lipid deposition that is common to dietary Cu levels.

In summary, we cloned the full-length sequences of five key genes (GRP78, PERK, IRE-1α and ATF-6α) involved in ER stress response from yellow catfish and investigated their molecular characterization, which will facilitate further studies on the regulation of ER stress response at the molecular level. mRNA expression patterns of these genes in different tissues also provided a basis
for further exploring the differential regulatory patterns of ER stress response in different tissues for the fish species. Dietary Cu excess activated ER stress and the UPR signaling in a tissue-specific manner in yellow catfish, which will help us to understand the effects of dietary Cu levels on the upstream pathway of lipid metabolism—the ER, and thus provide some novel insights on the nutrition of Cu in fish.

Acknowledgments

The authors thank the staff of Aquatic Animal Nutrition and Feed Laboratory of Huazhong Agricultural University for his assistance with excellent sample analysis. This work was supported by the National Natural Science Foundation of China (Grant no. 31422056), the Fundamental Research Funds for the Central Universities, China (Grant nos. 2662015PY017, 2014JQ002, 2013PY073), and by the PhD Candidate Research Innovation Project of Huazhong Agricultural University (Program No. 2014bs36).

LITERATURE CITED


(calreticulin) of skeletal muscle sarcoplasmic reticulum. J. Biol. Chem. 264: 21522-8.


Heat-shock responsive genes identified and validated in Atlantic cod (Gadus morhua) liver, head kidney and skeletal muscle using genomic techniques. BMC Genomics 11:72.


Lundebye, A.K., M.H.G. Berntssen, S.E.


Okada, K., T. Minamino, Y. Tsukamoto, Y. Liao, O. Tsukamoto et al., 2004 Prolonged endoplasmic reticulum stress in hypertrophic and failing heart after aortic constriction:
possible contribution of endoplasmic
reticulum stress to cardiac myocyte

Oyadomari, S., H.P. Harding, Y. Zhang,
M. Oyadomari, and D. Ron, 2008
Dephosphorylation of translation
initiation factor 2alpha enhances
glucose tolerance and attenuates
hepatosteatosis in mice. Cell Metab.
7: 520-532.

Ozcan, U., Q. Cao, E. Yilmaz, A.H. Lee,
N.N. Iwakoshi et al., 2004
Endoplasmic reticulum stress links
obesity, insulin action, and type 2

Palade, G., 1975 Intracellular aspects of
the process of protein synthesis.
Science 189: 347-358.

Penn, O., E. Privman, H. Ashkenazy, G.
Landan, D. Graur et al., 2010
GUIDANCE: a web server for
assessing alignment confidence
scores. Nucleic Acids Res. 38:
W23-W28.

Rebl, A., A. Brietzke, T. Goldammer, H.
M. Seyfert, 2014 GRP94 is encoded
by two differentially expressed genes
during development of rainbow trout
(Oncorhynchus mykiss). Fish Physiol.
Biochem. 40: 1917-1926.

Ron, D., and P. Walter, 2007 Signal
integration in the endoplasmic
reticulum unfolded protein response.

Rutkowski, D.T., J. Wu, and S.H. Backet, 
2008 UPR pathways combine to
prevent hepatic steatosis caused by
ER stressmediated suppression of
transcriptional master regulators. Dev.
Cell 15: 829-840.

Schröder, M., and R.J. Kaufman, 2005
The mammalian unfolded protein
response. Annu. Rev. Biochem. 74:
739-789.

Shaw, B.J., and R.D. Handy, 2006 Dietary
copper exposure and recovery in Nile
tilapia, Oreochromis niloticus. Aquat.
Toxicol. 76: 111-121.

Shiau, S.Y., and Y.C. Ning, 2003
Estimation of dietary copper


Xi, X.Z., and K.S. Ma, 2013 Molecular


Hu, 2013 GRP78 from grass carp (Ctenopharyngodon idella) provides cytoplasm protection against thermal and Pb\(^{2+}\) stress. Fish Shellfish Immunol. 34: 617-622.
Table 1 Nucleotide sequences of the primers used for the cDNAs cloning of GRP78, CRT, PERK, IRE-1α and ATF-6α genes

<table>
<thead>
<tr>
<th>Primers for partial fragment</th>
<th>Sequences (5′-3′)</th>
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</thead>
<tbody>
<tr>
<td><strong>GRP78-F</strong></td>
<td>AGAACACVGYTTYGATGCC</td>
</tr>
<tr>
<td><strong>GRP78-R</strong></td>
<td>TCCMACDGTCTCAATGCC</td>
</tr>
<tr>
<td><strong>CRT-F</strong></td>
<td>RGATGCGYTTYTATGC</td>
</tr>
<tr>
<td><strong>CRT-R</strong></td>
<td>GGYTTCCAYTCSCCCCTKRTA</td>
</tr>
<tr>
<td><strong>PERK-F</strong></td>
<td>AGTGYTTAGGTSGAGGCVG</td>
</tr>
<tr>
<td><strong>PERK-R</strong></td>
<td>GAGSAGTGARAGAGGGMGG</td>
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<tr>
<td><strong>IRE-1α-F</strong></td>
<td>ACTCTRACCTCTYTTTTG</td>
</tr>
<tr>
<td><strong>IRE-1α-R</strong></td>
<td>CCMCTRTCATTYACCTTCT</td>
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<tr>
<td><strong>ATF-6α-F</strong></td>
<td>TGGTMCTTGYTTTCTCGGC</td>
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<td><strong>ATF-6α-R</strong></td>
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<td>3′ GS-GRP78 01-F</td>
<td>TGAAGACTTTGACCACGCTG</td>
</tr>
<tr>
<td>3′ GS-GRP78 11-F</td>
<td>CTGGACAGGAGGATACAGGAA</td>
</tr>
<tr>
<td>3′ GS-CRT 01-F</td>
<td>GGATGATGAATACACCCACC</td>
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<td>3′ GS-CRT 11-F</td>
<td>GAGTGGAGAATGGGAGGAC</td>
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<td>3′ GS-PERK 01-F</td>
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<td>3′ GS-PERK 11-F</td>
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<tr>
<td>3′ GS-IRE-1α 01-F</td>
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<tr>
<td>3′ GS-IRE-1α 11-F</td>
<td>ACATCCCCTTTTCCACCT</td>
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<tr>
<td>3′ GS-ATF-6α 01-F</td>
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<td>3′ GS-ATF-6α 11-F</td>
<td>CTGGATCTTACTGCATCA</td>
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<tr>
<td>3′ RACE Outer</td>
<td>TACCGTCGTTTACTGTATTT</td>
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<td>3′ RACE Inner</td>
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<td>TTTCCAAAGGTAAGCCTCTG</td>
</tr>
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<td>5′ GS-CRT 01-R</td>
<td>TTACGTTTATGGCCAGGATG</td>
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<tr>
<td>5′ GS-CRT 11-R</td>
<td>CTCCATTCCATCCATCATC</td>
</tr>
<tr>
<td>5′ GS-PERK 01-R</td>
<td>CAGGGCTTTTGGGTATAG</td>
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<tr>
<td>5′ GS-PERK 11-R</td>
<td>TGTAAGCCCTCCAGGACATA</td>
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<tr>
<td>5′ GS-IRE-1α 01-R</td>
<td>CACCACCGGAGGATG</td>
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<tr>
<td>5′ GS-IRE-1α 11-R</td>
<td>TTTCGTCAGTCCCTCATTG</td>
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</table>
5′ GS-ATF-6α 01-R  TGAATGGGTCTCATAACAG
5′ GS-ATF-6α 11-R  AACTCTTCGCATCTCCCA
5′ RACE Outer    CATGGCTACATGCTGACAGCCTA
5′ RACE Inner     CGCGGATCCACAGCCTACTGATGATCAGTCGATG

Mixed bases: R-A/G; Y-C/T; M-A/C; K-G/T; S-G/C; W-A/T; H-A/T/C; B-G/T/C; V-G/A/C;

D-G/A/T.
Table 2 Feed formulation and proximate analysis of experimental diets (Chen et al. 2015).

<table>
<thead>
<tr>
<th>Ingredients (g kg⁻¹)</th>
<th>Adequate Cu</th>
<th>Cu deficiency</th>
<th>Cu excess</th>
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<tr>
<td>Casein</td>
<td>320</td>
<td>320</td>
<td>320</td>
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<tr>
<td>Gelatin</td>
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<td>80</td>
<td>80</td>
</tr>
<tr>
<td>Fish oil</td>
<td>25</td>
<td>25</td>
<td>25</td>
</tr>
<tr>
<td>Corn oil</td>
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<td>50</td>
<td>50</td>
</tr>
<tr>
<td>Wheat flour</td>
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<td>250</td>
<td>250</td>
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<td>Ascorbyl-2-polyphosphate</td>
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</tr>
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<td>NaCl</td>
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<td>10</td>
<td>10</td>
</tr>
<tr>
<td>NaH₂PO₄·2H₂O</td>
<td>10</td>
<td>10</td>
<td>10</td>
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<tr>
<td>CuSO₄·5H₂O</td>
<td>0.013</td>
<td>0</td>
<td>0.39</td>
</tr>
<tr>
<td>Vitamin premix</td>
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<td>5</td>
<td>5</td>
</tr>
<tr>
<td>Mineral premix (Cu-free)</td>
<td>5</td>
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<td>5</td>
</tr>
<tr>
<td>Betaine</td>
<td>10</td>
<td>10</td>
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<tr>
<td>Cellulose</td>
<td>224.987</td>
<td>225</td>
<td>224.61</td>
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</table>

Proximate analysis (% dry matter basis)

<table>
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<th></th>
<th>Adequate Cu</th>
<th>Cu deficiency</th>
<th>Cu excess</th>
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<tr>
<td>Moisture</td>
<td>8.63</td>
<td>8.58</td>
<td>8.92</td>
</tr>
<tr>
<td>Crude protein</td>
<td>34.38</td>
<td>34.21</td>
<td>34.29</td>
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<tr>
<td>Lipid</td>
<td>7.63</td>
<td>7.59</td>
<td>7.72</td>
</tr>
<tr>
<td>Cu (mg kg⁻¹)</td>
<td>4.18</td>
<td>0.76</td>
<td>92.45</td>
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Vitamin premix according to Tan et al. (2010); Mineral premix according to Tan et al. (2010) without Cu addition. CuSO₄·5H₂O(≥99.0% in purity): Sinopharm Chemical Reagent Co. Ltd., Shanghai, China.
<table>
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<th>Genes</th>
<th>GenBank accession No.</th>
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<th>Reverse primer (5'-3')</th>
<th>Size (bp)</th>
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<tr>
<td></td>
<td></td>
<td>CCCCCA</td>
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<td>CRT</td>
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<td>CGTGAAGAAGAG</td>
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<td>PERK</td>
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<td></td>
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<td>eIF2α</td>
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<td>AGGATGTGGTGAT</td>
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<tr>
<td>IRE-1α</td>
<td>KP687345</td>
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<td></td>
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<td>XBP-1</td>
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<td>GTGCTTCTCATTTC</td>
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<td>TTCATC</td>
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<td>ATF-6</td>
<td>KP687343</td>
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<td>GAPDH</td>
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Table 4 The information for GRP78, CRT, PERK, IRE-1α and ATF-6α cDNAs cloned from yellow catfish.

<table>
<thead>
<tr>
<th></th>
<th>GRP78</th>
<th>CRT</th>
<th>PERK</th>
<th>IRE-1α</th>
<th>ATF-6α</th>
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<td>Length (bp)</td>
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<td>1971</td>
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<td>Number of amino acids</td>
<td>652</td>
<td>418</td>
<td>1080</td>
<td>1038</td>
<td>657</td>
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Table 5 Amino acid sequence identity of GRP78, CRT, PERK, IRE-1α and ATF-6α between yellow catfish and other species (%). EMBL databases accession numbers: GRP78 (ENSDARG00000004665, ENSORLG0000000686, ENSXETG00000016838, ENSMGAG00000008510, ENSECAG00000024205, ENSMUSG00000026864, ENSG00000044574); CRT (ENSDARG00000043276, ENSORLG00000002923, ENSXETG00000007937, ENSMGAG00000002470, ENSECAG00000008164, ENSMUSG0000003814, ENSG00000179218); PERK (ENSDARG00000062139, ENSORLG00000017993, ENSXETG00000001926, ENSMGAG00000013189, ENSECAG00000020856, ENSMUSG00000031668, ENSG00000172071); IRE-1α (ENSDARG00000013997, ENSORLG00000013997, ENSXETG00000001926, ENSMGAG00000007831, ENSECAG00000011262, ENSMUSG00000020715, ENSG00000178607); ATF-6α (ENSDARG00000012656, ENSORLG00000016676, ENSXETG00000004747, ENSMGAG00000005185, ENSECAG00000022307, ENSMUSG00000026663, ENSG00000118217). The order of accession numbers of each gene corresponds to zebrafish, medaka, xenopus, turkey, horse, mouse and human, respectively.

<table>
<thead>
<tr>
<th></th>
<th>Zebrafish</th>
<th>Medaka</th>
<th>Xenopus</th>
<th>Turkey</th>
<th>Horse</th>
<th>Mouse</th>
<th>Human</th>
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<td>CRT</td>
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<td>PERK</td>
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</table>
Figure 1 (A) ClustalX alignment of the deduced amino acid sequence of GRP78 from *P. fulvidraco* and other species. The identical residues are shaded dark grey. Characteristic features are denoted as follows: SP (signal peptide) brace; C-terminal KDEL motif, an ER-retrieval sequence, asterisk; ATPase domain, dotted line; Substrate-binding domain, solid line. Duced amino acid sequences were obtained from *D. rerio* (GenBank/EMBL accession no. ENSDART00000010079) and *H. sapiens* (M19645). (B) The predicted peptide features of GRP78. SP, signal peptide.
**Figure 2** (A) ClustalX alignment of the deduced amino acid sequence of CRT from *P. fulvidraco* and other species. The identical residues are shaded dark grey. Characteristic features are denoted as follows: SP (signal peptide) brace; N-glycosylation site, triangle; C-terminal KDEL motif, an ER-retention sequence, asterisk; N domain, dashed line; P domain, solid line; C domain, dotted line; triplicate repeats A, solid lines box; triplicate repeats B, dotted lines box. Deduced amino acid sequences were obtained from *D. rerio* (GenBank accession no. NM_131047) and *H. sapiens* (NM_004343). (B) The predicted peptide features of CRT. SP, signal peptide.
Figure 3 (A) ClustalX alignment of the deduced amino acid sequence of PERK from *P. fulvidraco* and other species. The identical residues are shaded dark grey. Characteristic features are denoted thus: SP (signal peptide) brace; N-linked glycosylation site, ¥; the invariant lysine, ※; TM (transmembrane domain), solid lines box; Kinase (protein kinase), dotted lines box; stress sensing domain, solid line; effector domain, dotted line. Transmembrane regions were predicted from a hydropathy plot taking peaks with scores above 1.6 using a scan window size of 18 (Kyte and Doolittle, 1982). Deduced amino acid sequences were obtained from *M. musculus* (GenBank accession no. NM_010121) and *D. rerio* (XM_005156585). (B) The predicted peptide features of PERK. SP, signal peptide; TM, transmembrane domain; Kinase, kinase pretein.
Figure 4 (A) ClustalX alignment of the deduced amino acid sequence of IRE-1α from *P. fulvidraco* and other species. The identical residues are shaded dark grey. Characteristic features are denoted thus: SP (signal peptide) brace; TM (transmembrane domain), solid lines box; Kinase (protein kinase), dotted lines box; RNase (ribonuclease), bold dotted lines box; the conserved lysine in kinase domain II, ※; stress sensing domain, solid line; effector domain, dotted line. Transmembrane regions were predicted from a hydrophathy plot taking peaks with scores above 1.6 using a scan window size of 18 (Kyte and Doolittle, 1982). Deduced amino acid sequences were obtained from *M. musculus* (GenBank accession no. AF071777) and *O. latipes* (AB667982). (B) The predicted peptide features of IRE-1α. SP, signal peptide; TM, transmembrane domain; Kinase, protein kinase; RNase, ribonuclease.
Figure 5 (A) ClustalX alignment of the deduced amino acid sequence of ATF-6a from P. fulvidroco and other species. The identical residues are shaded dark grey. Characteristic features are denoted thus:

- **bZIP** (basic leucine zipper), dotted lines box; **TM** (transmembrane domain), solid lines box; **SIP**, (Site-1 cleavage site) triangle; **S2P**, (Site-2 cleavage site), asterisk; **stress sensing domain**, solid line; **effector domain**, dotted line. Transmembrane regions were predicted from a hydropathy plot taking peaks with scores above 1.6 using a scan window size of 18 (Kyte and Doolittle, 1982). Deduced amino acid sequences were obtained from H. sapiens (GenBank accession no. AB015856) and O. latipes (NM_001278901). (B) The predicted peptide features of ATF-6a. **bZIP**, basic leucine zipper; **TM**, transmembrane domain.
**Figure 6** Phylogenetic trees of based on the amino acid sequences of GRP78 (A), CRT (B), PERK (C), IRE-1α (D) and ATF-6α (E) from yellow catfish and the other vertebrate species, constructed by neighbor-joining method in MEGA 5.0 (Tamura et al., 2011) based on the JTT+G model (Jones et al., 1992) with 1000 bootstrap replicates. Accession numbers are shown next to each species. Numbers above branches indicate bootstrap support percentage over 50% in 1000 replicates.
Figure 7 qPCR analysis for GRP78, CRT, PERK, IRE-1α and ATF-6α across skeletal muscle (M), gill (G), head kidney (K), midintestine (I), brain (B), spleen (S), liver (L), heart (H), testis (T) and ovary (O). Data (mean±SEM, n=6) were normalized to housekeeping gene (β-actin and GAPDH) expressed as a ratio of the control (value at skeletal muscle = 1). Bars that do not share a common letter are significantly different among the different tissues (P < 0.05).

Figure 8 Effects of dietary Cu levels on the expression of GRP78, CRT, PERK, IRE-1α and ATF-6α mRNAs in the liver and muscle of yellow catfish after 8 weeks were quantified by qPCR. Data (mean±SEM, n = 3 replicate tanks, three fish were sampled for each tank) were normalized to housekeeping gene (β-actin and GAPDH) expressed as a ratio of the control (value at adequate Cu = 1). Different letters indicate significant differences among the treatments (P < 0.05).